Serbiluz

Invest Clin 60(1): 29 - 37, 2019 https://doi.org/10.22209/IC.v60n1a03

The impact of IL-10 gene polymorphism 1082A/G (rs1800896) on increased IL-10 secretion in patients with chronic kidney disease in the Kurdistan Region of Iraq.

Sarhang H. Azeez and Suhaila N. Darogha

JNIVERSIDAD

EL ZULIA

College of Education, Salahaddin University- Erbil, Iraq

Key words: chronic kidney disease; polymorphism; interleukin-10.

Abstract. Studies have indicated that interleukin-10 gene polymorphism at 1082A/G is associated with chronic kidney disease (CKD). The aim of this study was to determine II-10 gene polymorphism in CKD patients and identify the risk factors and prevalence of the disease among Kurdish patients. It was also aimed at finding out the serum levels of IL-10 in different genotypes, AA, GA and GG. The study included 108 patients with CKD: 54 on hemodialysis (HD) and 54 renal-transplanted (RT) and 54 healthy subjects. The mean age for HD, RT and healthy subjects was respectively 46.1, 36.8 and 40.2. Half of the HD patients and healthy subjects (50%) were male and half were female, while 55.6% of the RT patients were male and 44.4% female. According to the allele frequency of both G and A, there was no significant difference between both groups of patients and the healthy subjects (P=0.42). The AG genotype was independently associated with increased risk of CKD undergoing HD and RT, while the GG genotype showed an increased risk for renal failure. The levels of serum IL-10 concentrations increased significantly in both groups of patients compared to the healthy subjects. Regarding the genotypes, the genotype AA had the highest concentration among the patients, while a high level was found in genotype GA in the healthy subjects. The lowest level of this cytokine was found in both genotypes GG and AA in the healthy subjects. The findings of the present study revealed that IL-10 gene polymorphism at 1082 A/G (rs1800896) and increased concentration of serum IL-10 were associated with increasing chronic kidney disease in the Iraqi Kurdish population.

Corresponding author: Sarhang H. Azeez, College of Education, Salahaddin University- Erbil, Iraq. Phone: +964 750 462 4662. Email: Sarhang.azeez@su.edu.krd

Impacto del polimorfismo 1082A/G (rs1800896) del gen de IL-10 sobre la secreción de IL-10 en pacientes con enfermedad renal crónica en la región Kurdistán de Irak.

Invest Clin 2019; 60 (1): 29-37

Palabras clave: enfermedad renal crónica; polimorfismo; interleucina-10.

Resumen. Varios estudios han indicado que el polimorfismo 1082A/G (rs1800896) del gen de IL-10 está asociado con la enfermedad renal crónica (ERC). El propósito de este estudio fue determinar el polimorfismo del gen de la IL-10 en pacientes con ERC e identificar los factores de riesgo y prevalencia de la enfermedad entre pacientes Kurdos. También se determinaron las concentraciones séricas de IL-10 en los diferentes genotipos AA, GA y GG presentes. El estudio incluyó 108 pacientes con ERC: 54 en hemodiálisis (HD) y 54 con trasplante renal (TR) y en 54 sujetos sanos. Las edades promedio para los HD, TR y sujetos sanos fueron 46,1, 36,8 y 40,2 años, respectivamente. La mitad de los pacientes en HD y de los sujetos sanos eran hombres y la otra mitad eran mujeres, mientras que el 55,6% de los pacientes con TR eran hombres y 44,4 % eran mujeres. Considerando la frecuencia de los alelos G v A, no hubo diferencias estadísticamente significativas entre ambos grupos de pacientes y los sujetos sanos (p = 0.42). El genotipo AG se encontró independientemente asociado con un riesgo aumentado de pacientes con ERC bajo HD y TR, mientras que el genotipo GG mostró un riesgo mayor de falla renal. Las concentraciones séricas de IL-10 se encontraron aumentadas significativamente en ambos grupos de pacientes, comparadas con las de los sujetos sanos. En consideración a los genotipos, el genotipo AA tenía las más altas concentraciones entre los pacientes, mientras que se encontró un mayor nivel en el genotipo GA en los sujetos sanos. Las concentraciones más bajas de esta citocina se encontraron en los genotipos GG v AA de los sujetos sanos. Los hallazos del presente estudio revelaron que el polimorfismo 1082A/G (rs1800896) del gen de la IL-10 y las concentraciones séricas aumentadas de IL-10 estaban asociadas con aumentos de la enfermedad renal crónica en la población Kurda de Iraq.

Recibido 30-10-2018 Aceptado 22-01-2019

INTRODUCTION

Progressive kidney dysfunction is known as CKD. The HD, peritoneal dialysis and RT, which are renal replacement therapies, may extend survival in patients with end-stage-renal-disease (ESRD) and provide a good quality of life in most cases (1). In HD patients, the levels of several cytokines, such as IL-10, IL-2, IL-4, IL-5 and IFN-γ, increase (2). Morbidity and mortality, which are independent risk factors in CKD patients, remain unchanged despite undoubted improvements in haemodialysation techniques, and are due to high levels of interleukins, the presence of metabolic acidosis, chronic inflammation, malnutrition, anemia, and cardiovascular disease (CVD) (3).

Furthermore, ESRD is associated with impaired cellular and humoral immunity

and persistent immune system activation (4,5). Large amounts of pro and anti-inflammatory cytokines produced by circulating monocytes and regulatory T cells (CD4+/CD25+) lead to limitation of inflammatory activation and subsequent pathogen elimination (6).

Because of endothelium function, atherosclerosis is accelerated directly by proinflammatory cytokines during early stages of CKD 3 and 4 (7-9). Constant inflammatory state, during which the patient suffers from its nutritional state, suppresses bone marrow stem cells leading to anemia, and shares essentially in erythropoietin therapy resistance, is correlated with poor outcomes of dialysis (10-13). Development of immune tolerance and pro-inflammatory cytokines reduced by the activity of IL-10 leads to a decrease in recipient T-cell activation and allogenic responses (14).

Individuals with adequate IL-10 production during ESRD have a better control over infection and uremia-associated state and experience less coronary artery disease (15-18). There are multiple polymorphisms in IL-10 gene promoter especially at positions _1082 (rs1800896), _819 (rs1800871), and _ 592 (rs1800872) which intensely affect the IL-10 levels (19-21).

The presence of essential biological impacts on transcriptional activity results in high-, intermediate-, and low-producing phenotypes of IL-10 gene promoter polymorphism. IL-10 high producers show better immune response and infection control and lower risk of cardiovascular death (22).

Cytokine production and expression are genetically determined; therefore, allelic variants of cytokine genes arising from nucleotide polymorphisms within regulatory region have been described in the last two decades (23).

MATERIALS AND METHODS

Patients and control group

The required samples were collected from January to November 2017. In so doing,

54 patients on maintenance hemodialysis at the Hawler Dialysis Centre and 54 patients on renal replacement therapy from private clinics at Hawler city were chosen for this study. Fifty-four healthy subjects were also selected as a control group, who were homogeneous with the patient groups regarding their age and gender. Clinical and laboratory data were obtained from the subjects, and a questionnaire was filled up for each patient and healthy individual in this study.

Blood samples

Peripheral blood samples (5 mL) were obtained from the groups and placed into two different tubes. About 2 mL of the blood samples were placed in heparinized tubes for DNA extraction from the whole blood or mononuclear cells, and the tubes were stored at -70°C until genomic extraction was obtained. The remaining 3 mL were placed in a gel tube to obtain the serum. The tubes were centrifuged at 10000 rpm for 10 minutes, and the serum was put into 1.5 ml Eppendorf tube and stored at -70°C until cytokine level estimation was carried out.

Determination of serum IL-10

Levels of serum IL-10 pg/mL were determined by enzyme linked immune sorbent assay (ELISA), and the Cloud Clone Corp. kit was used according to the manufacturer's protocol.

Cytokine genotyping

Using the manufacturer's instruction for QIAmp DNA mini kit (Qiagen, Hilden, Germany), peripheral blood mononuclear cells were used to obtain genomic DNA for both patients and control groups. For the IL-10 genotype at (-1082 G/A) (rs1800896), the amplification refractory mutational system method (ARMS-PCR) was utilized. The assays were performed in a 20 μ L reaction volume containing 40 ng genomic DNA, 1.5 mM dNTPs, 25 mM MgCl₂, 1 μ L of 10 pmol each primer and 0.4 units of Taq polymerase (Fermentas, Maryland, USA) in 1X Reaction Buffer. The primer sequences were as follows: IL-10 generic primer, 5'-CAGTGC-CAACTGAGAATTTGG-3', IL-10 (G) Allele Primer 5'-CTACTAAGGCTTCTTTGGGAG-3, and IL-10 (A) Allele Primer 5'-ACTACTA-AGGCTTCTTTGGGAA-3. The PCR reaction was carried out in a thermal cycler (PX2) with the following cycling conditions: 95°C for 3 minutes, followed by 35 cycles at 95°C for 45 seconds, 58°C for 40 seconds, 72°C for 1 minute, and finally a 7-minute extension at 72°C. The amplicon size was 254bp. The amplified products were analyzed on 2% agarose gel.

Statistical Analysis

All statistical analyses were performed using SPSS 19.0 (SPSS Inc., Chicago, IL, USA). Normally distributed variables were expressed as mean \pm SD as appropriate. The level of statistical significance was set at P<0.05. Intergroup comparisons were assessed for categorical variables and serum cytokine concentration using ANOVA. For IL-10 gene polymorphism 1082A/G (rs1800896), allele was counted by direct allele counting. The Hardy–Weinberg equilibrium was assessed with the chi square (X^2)-test. Descriptive data were presented as mean \pm standard deviation (SD). Genotype and allele frequencies were compared between the groups using a (X^2)-test of independence with 2 x 2 contingency tables and z statistics. Statistical significance of the variables was established at the level P<0.05.

RESULTS

In the present study, CKD affected both young and old subjects. Both genders were equally influenced by this disease, too. The mean age in the patient groups with HD (n=54) and RT (n=54) were 46.1 and 36.8 years, respectively. With regard to the subjects' gender, 50% of the HD patients were males and 50% female, while 55.6% of the RT patients were males and the rest 44.4% were females. In this regard, there was no significant differences between the patient groups and the control group (p>0.05). The results also revealed that 27.8% and 20.4% of the patients respectively in HD and RT groups smoked. However, about 26% of the HD patients and 13% of the RT patients had diabetes. Moreover, hypertension was observed in about 50% and 16.7% of the HD and RT patients, respectively. It was also seen that 20.4% and 24.1% of the HD and RT patients had a family history of these conditions, respectively (Table I).

Groups Characters	Hemodialysis N=54	Renal Transplanted N=54	Control N=54
Age (Mean±SD)	46.1±1.6	36.8±2.8	40.2±1.9
Sex:			
Male	27/54(50%)	30/54(55.6%)	27/54(50%)
Female	27/54(50%)	24/54(44.4%)	27/54(50%)
BMI (Mean±SD)	24.83 ± 0.5	24.17 ± 1.0	23.62 ± 0.45
Smokers	15/54(27.8%)	11/54(20.4%)	
Diabetes	14/54(26%)	7/54(13%)	
Hypertension	25/54(50%)	9/54(16.7%)	
Family History	11/54(20.4%)	13/54(24.1%)	

 TABLE I

 THE DEMOGRAPHIC DISTRIBUTION OF THE STUDIED GROUPS

Detection of IL-10 gene polymorphism 1082A/G (rs1800896)

Genotypic frequencies of alleles showed different band patterns of amplified fragments for all of the HD, RT and healthy subjects (Table II). For IL-10 gene polymorphism1080, the results were as follows: GG: 30 (55.6%), 20* (37%) and 15* (27.8%) respectively for healthy subjects, HD and RT patients, while the distributions of GA were 20 (37%), 31* (57.4%) and 30 (55.6%) for healthy subjects, HD and RT patients, respectively. However, the distribution for AA was as follows: healthy subjects 4 (7.4%), hemodialysis 3 (5.6%), and renal transplanted patients 9 (16.6%). The results of the present study showed a significant difference between healthy subjects and both patient groups in terms of GG distribution (P<0.05). Also, healthy subjects and HD patients were significantly different regarding GA distribution (P<0.05). The Chi-square (X^2) result for the HD patients was 4.067 with a p-value

of 0.047, which was consistent with HWE. However, the X^2 results for the RT patients and healthy subjects were 0.843 and 0.06, respectively (p>0.05), which was not consistent with HWE. Furthermore, the statistical analysis of genotypes and allele numbers was confirmed by direct allele counting and Chi-square test.

The results of the current study showed that the levels of serum IL-10 concentrations significantly increased in both groups of patients compared to the healthy subjects (P<0.0001). Regarding different genotypes and the concentration of serum IL-10, data analysis showed a significant increase in the patients compared to the healthy subjects (p<0.0001) for genotype GG, p<0.001 for genotype GA, and p < 0.01 for genotype AA). Genotype GG had the highest concentration among the patients, while the highest level was found in genotype GA in the healthy subjects. The lowest level of this cytokine was found in genotype AA in the healthy subjects (Table III and Figs. 1 and 2).

IL-10 1082 G/A (rs1800896)	Hemodialysis	Renal Transplanted	Healthy Subjects	р
No. of genotypes	N=54	N=54	N=54	
GG	20* (37%)	15* (27.8%)	30 (55.6%)	p<0.05
GA	31* (57.4%)	30 (55.6%)	20 (37%)	p<0.05
AA	3 (5.6%)	9 (16.6%)	4 (7.4%)	p>0.05
Allele frequency				
G	0.66	0.56	0.74	
А	0.34	0.44	0.26	
X^2	4.067	0.843	0.060	

 TABLE II

 IL-10 GENOTYPE AND ALLELE FREQUENCY OF STUDIED GROUPS

* Significant difference.

Cytokine	e	Hemodialysis (n=54)	Renal Transplanted (n=54)	Healthy Subjects (n=54)	р				
IL-10 (pg/mL)		$26.29^{a} \pm 2.422$	$24.41^{\rm b} \pm 1.559$	15.00 ± 0.918	p<0.0001				
IL-10 (pg/mL)	GG	$29.79^{a} \pm 1.45$	25.85 ± 2.32	14.63 ± 1.392	p<0.0001				
	GA	$26.28^{a} \pm 3.173$	23.72 ± 2.037	15.00 ± 0.918	p<0.001				
	AA	$25.43^{a} \pm 1.929$	22.10 ± 3.306	14.38 ± 0.963	p<0.01				

 TABLE III

 CONCENTRATION OF SERUM IL-10 pg/mL (Mean±SD) IN STUDIED GROUPS

 REGARDING THEIR GENOTYPES

^a: represents the significant difference between Hemodialysis and control group.

^b: represents the significant difference between Renal transplanted and control group.

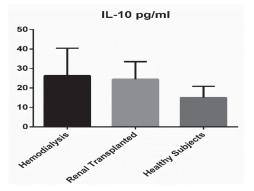


Fig. 1. Concentration of IL-10 pg/mL in the patients and healthy subjects.

As indicated in Table III above, intergroup comparisons indicated that the patient groups had a significantly higher level of the studied cytokine (i.e. IL-10) than the control group. However, intragroup comparison between the levels of different genotypes in each group revealed that the levels of the studied genotypes were not significantly different in any of the studied groups. This finding is clearly understandable by comparing the data presented in Table II above and Fig. 2 below.

DISCUSION

The results of the present study are in agreement with those of the study carried out by Zhang *et al* (24) who reported that IL-6 and IL-10 levels increased in HD patients. Another study showed a high level

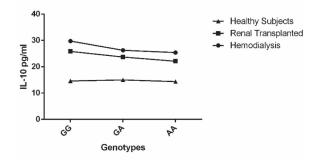


Fig. 2. Level of IL-10 according to the genotypes of studied groups.

of IL-10 in CKD (HD) in comparison with control individuals, and a higher IL-10 expression in long-term HD patients compared with short-term HD ones (25).

Research has revealed that the serum level of IL-10 increases as a result of uraemic monocytes and that kidney clearance drops in ESRD (26). IL-10 genotype is considered as a risk factor for both ESRD and CVD (6, 27,28). High level of IL-10 was found in G allele at position -1082 (rs1800896) promoter region (29).

Although regular HD causes decreased levels of mortality in ESRD patients, it is considered a condition associated with inflammation (30). In CKD the variation in concentrations of both pro-inflammatory and anti-inflammatory cytokines were several times higher than healthy subjects, this is due to both decreased renal clearance and increased production of cytokines (31,32).

The cytokine secretion varied between different individuals in this study due to different levels of their response to the stimuli. This alteration is determined by different promoter regions of polymorphism, which may lead to a person's susceptibility to a variety of chronic inflammation induced by cytokines.

Similar to the present study, there was a strong association between nucleotide polymorphism of IL-10 and functions of kidneys during CKD. Despite the patients' age, gender, and body mass index, the association remained stable, and polymorphism is reported as a risk factor for CKD outcomes (33). It is suggested that IL-10 production is determined genetically and can be controlled at the transcriptional level (34).

Research has indicated that the -1082 G allele is associated with higher IL-10 production, while the A allele with lower IL-10 production. These results confirm the findings of the present study (35). There is a higher IL-10 production in G/G genotype compared to other genotypes, which has been confirmed by previous studies. The susceptibility and/or severity of the disease by alteration in levels of both pro- and antiinflammatory cytokines is genetically determined by lower IL-10 production (34).

In genotype combinations, there might be a strong regulatory increase in IL-10 production on the status of patients during CKD. Indeed, the results of the current study demonstrated a gradation of risk in the relationship between outcomes and genotype combinations that would be expected to predispose to a pro-inflammatory state with a consistent linear trend demonstrable on univariate analysis (36).

The results of the present study indicated that IL-10 serum level was genetically determined. They also showed differences between various genotypes in IL-10 gene polymorphism at the 1082 region (rs1800896) in CKD patients including HD and RT. The prevalence of the CKD was in the age group near 40 years. According to the results of this study, both genders were equally susceptible to CKD. It was also concluded that hypertension and diabetes were complications or risk factors for the development of the disease.

35

ACKNOWLEDGMENTS

The authors would like to thank all of the participants of this study. Grateful thanks also go to all of the nurses who helped with conduction of this study.

REFERENCES

- 1. Pedro T, Sara A, Combs, and Isaac T. Peritoneal dialysis: update on patient survival. Clin Nephrol 2015; 83: 1-10.
- Rios D, Pinheiro M, de Oliveira Junior W, Braga K, Carvalho A, Martins-Filho O, Simões E, Silva AC, Dusse LMS. Cytokine signature in end-stage renal disease patients on hemodialysis. Dis Markers 2017; 9678391.
- **3.** Zahed NS and Chehrazi S. The evaluation of the relationship between serum levels of interleukin-6 and interleukin-10 and metabolic acidosis in hemodialysis patients. Saudi J Kidney Dis Transpl 2017; 28(1):23-29.
- 4. Daichou Y, Kurashige S, Hashimoto S, Suzuki S. Characteristic cytokine products of Th1 and Th2 cells in hemodialysis patients. Nephron 1999; 83: 237-245.
- 5. Ando M, Shibuya A, Yasuda M, Azuma N, Tsuchiya K, Akiba T, Nitta K. Impairment of innate cellular response to in vitro stimuli in patients on continuous ambulatory peritoneal dialysis. Nephrol Dial Transplant 2005; 20: 2497-2503.
- 6. Girndt M, Ulrich C, Kaul H, Sester U, Sester M, Kohler H. Uremia-associated immune defect: The IL-10- CRP axis. Kidney Int Suppl 2003; 84: 76-79.
- Menon V, Greene T, Wang X, Pereira A, Marcovina S, Beck G, Kusek JW, Collins AJ, Levey AS, Sarnak MJ. C-reactive protein and albumin as predictors of all-cause and cardiovascular mortality in chronic kidney disease. Kidney Int 2005; 68: 766-772.

- 8. Honda H, Qureshi AR, Heimburger O, Barany P, Wang K, Pecoits-Filho R, Stenvinkel P, Lindholm B. Serum albumin, Creactive protein, interleukin 6, and fetuin A as predictors of malnutrition, cardiovascular disease, and mortality in patients with ESRD. Am J Kidney Dis 2006; 47: 139-148.
- **9. Stenvinkel P.** Inflammation in end-stage renal disease: the hidden enemy. Nephrology Carlton 2006a; 11: 36-41.
- 10. Cooper AC, Mikhail A, Lethbridge MW, Kemeny DM, Macdougall IC. Increased expression of erythropoiesis cytokines (IFN- γ , TNF- α , IL-10 and IL-13) by T cells in patients exhibiting a poor response to erythropoietin therapy. J Am Soc Nephrol 2003; 14: 1776-1784.
- 11. Kalantar-Zadeh K, McAllister CJ, Lehn RS, Lee GH, Nissenson AR, Kopple JD. Effect of malnutrition- inflammation complex syndrome on EPO hyporesponsiveness in maintenance hemodialysis patients. Am J Kidney Dis 2003; 42: 761-773.
- **12.** Stenvinkel P. New insights on inflammation in CKD genetic and non-genetic factors. Nephrol Ther 2006; 2: 111-119.
- **13.** De Francisco A, Stenvinkel P, Vaulont S. Inflammation and its impact on anaemia in chronic kidney disease: from haemoglobin variability to hyporesponsiveness. NDT Plus 2009; 2 (1):18-26.
- 14. Luo X, Miller S, Shea L. Immune tolerance for autoimmune disease and cell transplantation. Ann Rev Biomed Eng 2016; 18: 181–205.
- **15.** Girndt M. Humoral immune responses in uremia and the role of IL-10. Blood Purif 2002; 20: 485-488.
- 16. Seyrek N, Karayaylali I, Balal M, Paydas S, Aikimbaev K, Cetiner S, Seydaoglu G. Is there any relationship between serum levels of interleukin-10 and atherosclerosis in hemodialysis patients. Scand J Urol Nephrol 2005; 39: 405-409.
- 17. Girndt M, Heine GH, Kohler H, Dial Gene Consortium. Gene polymorphism association studies in dialysis: Anemia and host immunity. Semin Dial 2006; 19: 227-31.
- 18. Litjens N, Huisman M, van den Dorpel M, Betjes M. Impaired immune responses and antigen-specific memory CD4+ T cells in haemodialysis patients. J Am Soc Nephrol 2008; 19: 1483-1490.

- 19. Crawley E, Kay R, Sillibourne J, Patel P, Hutchinson I, Woo P. Polymorphic haplotypes of the interleukin-10 5_ flanking region determine variable interleukin- 10 transcription and are associated with particular phenotypes of juvenile rheumatoid arthritis. Arth Rheum 1999; 42 (6):1101– 1108.
- 20. Mäurer M, Kruse N, Giess R, Toyka K, Riekmann P. Genetic variation at position-1082 of the interleukin 10 (IL10) promotor and the outcome of multiple sclerosis. J Neuroimmunol 2000; 104(1):98-100.
- 21. Moudi B, Heidari Z, Mahmoudzadeh-Sagheb H, Hashemi M, Metanat M, Khosravi S, Farrokh P. Association between IL-10 gene promoter polymorphisms (-592 A/C, -819 T/C, -1082 A/G) and susceptibility to HBV infection in an Iranian population. Hepatitis Monthly 2016; 16(2): e32427.
- 22. Girndt M, Kaul H, Sester U, Ulrich C, Sester M, Georg T, Kohler H. Anti-inflammatory interleukin-10 genotype protects dialysis patients from cardiovascular events. Kidney Int 2002; 62: 949-955.
- 23. Wilson AG, DE Vries N, Pociot F, di Giovine FS, van der Putte LB, Duff GW. An allelic polymorphism within the human tumor necrosis factor-alpha promoter region is strongly associated with HLA A1, B8, and DR3 alleles. J Exp Med 1993; 177:557–560.
- 24. Zhang W, Wang W, Yu H, Zhang Y, Dai Y, Ning C, Tao L, Sun H, Kellems RE, Blackburn MR, Xia Y. Interleukin 6 underlies angiotensin II-induced hypertension and chronic renal damage. Hypertension 2012; 59:136-44.
- 25. Brunet P, Capo C, Dellacasagrande J, Thirion X, Mege JL, Berland Y. IL-10 synthesis and secretion by peripheral blood mononuclear cells in haemodialysis patients. Nephrol Dial Transplant 1998; 13:1745-51.
- 26. Stenvinkel P, Ketteler M, Johnson R, Lindholm B, Pecoits-Filho R, Riella M. IL-10, IL-6, and TNF-α: Central factors in the altered cytokine network of uremia: The good, the bad, and the ugly. Kidney Int 2005; 67: 1216-1233.
- 27. Loppnow H, Werdan K, Buerke M. Vascular cells contribute to atherosclerosis by cyto-

kine- and innate-immunity-related inflammatory mechanisms. Innate Immun 2008; 14: 63-87.

- 28. Han X, Kitamoto S, Lian Q, Boisvert WA. Interleukin-10 facilitates both cholesterol uptake and efflux in macrophages. J Biol Chem 2009; 284: 32950-32958.
- 29. Liu Z, Guo J, Wang Y, Li K, Wei Y, Sun Q, Xu Q, Xu C, Yan X, Tang B. Lack of association between IL-10 and IL-18 gene promoter polymorphisms and Parkinson's disease with cognitive impairment in a Chinese population. Scientific Reports 2016; 6:19021.
- **30.** Ori Y, Bergman M, Bessler H, Zingerman B, Levy-Drummer R, Gafter U, Salman H. Cytokine secretion and markers of inflammation in relation to acidosis among chronic hemodialysis patients. Blood Purif 2013; 35:181-6.
- 31. Kimmel PL, Phillips TM, Simmens SJ, Peterson RA, Weihs KL, Alleyne S, Cruz I, Yanovski JA, Veis JH. Immunologic function and survival in hemodialysis patients. Kidney Int 1998; 54:236-44.
- 32. Castillo-Rodríguez E, Pizarro-Sánchez S, Sanz AB, Ramos AM, Sanchez-Niño MD, Martin-Cleary C, Fernandez-Fernandez B, Ortiz A. Inflammatory cytokines as uremic toxins: "ni son todos los que están, ni están todos los que son". Toxins 2017: 9(4)114.

33. Sinuani I, Beberashvili I, Averbukh Z and Sandbank J. Role of IL-10 in the progression of kidney disease. W J Trans 2013; 3(4): 91–98.

37

- **34.** Iyer S, and Cheng G. Role of Interleukin 10 Transcriptional regulation in inflammation and autoimmune disease. Crit Rev Immunol 2012; 32(1): 23–63.
- **35.** Guo J, He Y, Chen F, Jiang M, Gao S, Su Y. The A to G polymorphism at -1082 of the interleukin-10 gene is rare in the Han Chinese population. Mol Med Rep 2012; 6: 894-896.
- 36. Vaidyanathapuram S, Balakrishnan, Daqing G, Madhumathi R, Bertrand L, Jaber H. Cytokine gene polymorphisms in hemodialysis patients: Association with comorbidity, functionality, and serum albumin. Kidney Int 2004; 65: 1449–1460.